amount of ATP and is independent of CoA, thus suggesting that succinyl phosphate may be formed, (d) in experiments with sulfanilamide,<sup>10</sup> a CoAand ATP-dependent decrease in free amide is observed; this decrease is greater if carbon dioxide production is inhibited, suggesting that succinyl-CoA, as well as succinyl phosphate, may be formed. A detailed treatment of the experimental results and of the postulated mechanism of succinate activation will be presented elsewhere.

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(10) D. R. Sanadi and J. W. Littlefield, J. Biol. Chem., 193, 683 (1951).

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## SULFATED NITROGENOUS POLYSACCHARIDES AND THEIR ANTICOAGULANT ACTIVITY<sup>1</sup>

Sir:

N-Deacetylated chitin, previously swollen with pyridine, was heterogeneously sulfated at 100° for 1 hour with chlorosulfonic acid and pyridine to yield a product, isolated (inorganic salts were removed by dialysis) as the amorphous, water-soluble sodium salt, containing essentially two N-sulfate and one O-sulfate groups per anhydrodisaccharide unit;  $[\alpha]^{25}D - 23^{\circ}$  (c 1.5, water). Anal. Calcd. for  $C_{12}H_{19}O_7(NSO_3Na)_2(OSO_3Na): C, 22.93; H, 3.05; N,$ 4.46; S, 15.31; Na, 10.98. Found: C, 22.68; H, 3.08; N, 4.02; S, 15.6; Na, 11.2; -NH<sub>2</sub> (by ninhydrin), absent; NAc, absent. This preparation exhibited the behavior in the Van Slyke amino acid assay characteristic of the acid-labile N-sulfate group present in heparin.<sup>2,3</sup> Its anticoagulant activity was 56 International Units (I. U.)/mg. The animal (mouse intravenous) toxicity was approximately double that of heparin, a finding believed to be due to the unsuitably high molecular size of the substance.

(1) Supported by the Bristol Laboratories, Inc., Syracuse, N. Y., (R. F. Project 432).

(2) M. L. Wolfrom and W. H. McNeely, This JOURNAL. 67, 748 (1945).

(3) J. E. Jorpes, H. Boström and V. Mutt, J. Biol. Chem., 183, 607 (1950).

Chondroitinsulfuric acid (from cartilage) was essentially homogeneously N-deacetylated with 45% NaOH (25, 48 hr.) under nitrogen and in the presence of antioxidants (benzyl alcohol and sodium sulfite) and was sulfated as described above (but at  $80-90^{\circ}$ ). The product was isolated as the amorphous sodium salt and is under further analytical characterization; -NH<sub>2</sub> (by ninhydrin), absent. The anticoagulant activity was 48 I. U./mg. The same sample of sodium chondroitin sulfate was subjected to the above sulfation procedure without preliminary N-deacetylation and the product, isolated in the same manner, showed an anticoagulant activity of ca. 10 I. U./mg. Sodium heparinate was re-sulfated under these conditions with a reduction in its activity from 110 to 55 I. U./mg. and an increase in the sulfur content from 12 (initial) to 14.4%

Methyl 2-amino-2-deoxy-*β*-D-glucopyranoside hydrochloride<sup>4</sup> was sulfated as above to produce the amorphous, water-soluble barium salt of the Nsulfate, tri-O-sulfate;  $[\alpha]^{25}D + 4^{\circ}$  (c 3.4, H<sub>2</sub>O). Anal. Calcd. for  $C_7H_{11}NO_{17}S_4Ba_2 \cdot 2H_2O$ : S, 15.63; Ba, 33.48. Found: S, 15.64; Ba, 32.98. A 3  $\times$  $10^{-4}$  M solution of this substance in 0.004 N HCl at 95° lost 1.0 mole of sulfate in  $\leq 20$  min. with the concomitant release of the free amino group (ninhydrin). The O-sulfate was removed relatively more slowly and only completely so after 12 hr. Previously reported results<sup>2</sup> on the inactivation of heparin by mild acidity were considered to involve a negligible sulfate loss. On the basis of our present knowledge of the heparin molecule,<sup>6</sup> this sulfate loss is about equivalent to the amino group released so that a sulfate group shift<sup>5</sup> is not a required postulation.

The above results show that the sulfamic acid group is a potent contributor to anticoagulant activity. Experiments are now underway to determine the optimum molecular size for these chemically modified polysaccharides.

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<sup>(4)</sup> J. C. Irvine, D. McNicoll and A. Hynd, J. Chem. Soc., 99, 250 (1911).

<sup>(5)</sup> M. L. Wolfrom, R. Montgomery, J. V. Karabinose and P. Rathgeb. This JOURNAL, 72, 5796 (1950).